

Protoporphyrin IX Associated with Visible Light for the Treatment of *Trichophyton rubrum* Causing Onychomycosis - An Updated Review

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Abstract

*Systemic medications used during the treatment of onychomycosis caused by *Trichophyton rubrum* may have relapse, making them costly and insignificant for the patient. Photodynamic therapy (PDT) is an advantageous therapeutic option for disease control, mainly due to the absence of risk of microbial resistance. The action of PDT is combined by three elements: photosensitizer (PS), visible light (VL) and molecular oxygen, leading to the formation of reactive oxygen species (ROS) or singlet oxygen formation (¹O₂), both ROS (type I mechanism) as ¹O₂ (type II mechanism) induce damage and death to microbial cells. This research was proposed as a study through a non-systematic review, to investigate the action of PS "Protoporphyrin IX" (Pp IX) associated with visible light on *T. rubrum*, contemplating clinical and relevant data regarding the treatment of onychomycosis by PDT. PubMed survey was conducted from June 2021 to April 2022. The research strategy included clinical trials, randomized trials, systematic reviews, meta-analyses and reviews in English. Pp IX has affinity for *T. rubrum*, which is justified by the incorporation of Pp IX into the cell membrane, which led to inhibition by LV irradiation. Thus, the photodynamic process of Pp IX may lead to cell death by type I and II mechanisms. The data found are promising, however, new studies in vitro and in vivo are suggested, since few studies have been found related to the theme.*

Keywords: *Trichophyton rubrum*; Onychomycosis; Phototherapy; Protoporphyrin IX; Low-level light therapy; Oxygen-reactive species; Singlet oxygen; Drug Induced Liver Injury; Roussel Uclaf Causality Assessment Method

1. Introduction

Onychomycosis, known as nail had, is an infection caused by dermatophytes, non-dermatophyte fungi (NDF) or yeasts. In particular, this disease is mainly affected by dermatophytes (90%), with *Trichophyton rubrum* as the main causative agent (71%). It usually presents with a white or yellowish-brown discoloration of the nail, onycholysis and/or thickening of the nail plate^[1-4]. Due to these aspects, onychomycosis brings an aesthetic problem to the patient, consequently, ends up distancing itself from society and with low self-esteem affecting quality of life.

The contagion of the disease occurs by direct contact of the fungus with the nail through the distal subungueal area and/or the lateral nail plate (LNP), this means that the diseased nail can contaminate other healthy nails. This can be explained by the fact that the nail unit does not have effective cellular immunity, and is therefore susceptible to fungal infection^[5]. In addition, fungal production of enzymes with proteolytic, keratinolytic and lipolytic activities helps to degrade keratin in the nail plate and this facilitates fungal invasion of the nail, increasing the risk of infection^[6,7]. For Peres *et al.*^[8], infections caused especially by dermatophyte *T. rubrum*, is related to the failure of the host's immune response or the ability of the pathogen to escape the defense system and thus remain in the superficial layers of the skin. It should also be added that the components of the fungal cell wall, such as galactomannans scans, may have immunosuppressive effects, and membrane transport proteins, important virulence factors that guarantee the success of colonization in favorable environments^[9,10].

The growth mechanism of dermatophytes is related to keratinized tissue and nail pH. Fungi grow very well in a medium containing rigid keratin and are able to use this tissue as the only source of carbon and nitrogen. At alkaline pH, two subtilisins (Sub3 and Sub4) and metalloproteases of the family of fungalisins (family M36) are the main endoproteases secreted by dermatophytes, together with exopeptidases. The latter enzyme catalyzes the cleavage of the terminal peptide bond by releasing two aminopeptidase leucines from the M28 family (Lap1 and Lap2) and two dipeptidil peptidases from the S9 family (DppIV and DppV). In a medium with acid pH, dermatophytes secrete an aspartic protease of the peptin family (Pep1) as an endoprotease and tryptophytin of the sedolysine family, X-proline peptidases and carboxypeptidases of the Family S10 as exoproteases^[11,12,13]. These considerations point out that dermatophyte enzymes have the ability to hydrolyse keratin and survive in protein medium and are still resistant to acid and alkaline pH.

Conventional treatment of nail had involves topical or oral therapy. Topical treatment is more compatible with the patient, being devoid of adverse effects, as long as the infections are mild, but suffers a setback, which is inadequate penetration into the nail^[14]. This factor can be explained by the formation of fungal biofilms, allowing fungi to avoid therapies and contribute to antifungal resistance^[15]. In the case of persistent infection, oral treatment is used, although quite effective, they are hepatotoxic and cause drug interactions. It is noteworthy that these antifungal therapies often result in failure and commonly used antifungals have limited cellular targets. Resistant strains also appear, concomitantly with high-cost

treatments for long periods, with probable side effects, in addition to the recurrence of the disease after discontinuation of therapy^[8,14,16].

The difficulties in the treatment of fungal infections by conventional methods have led to the search for a safer, more effective, easy-to-perform and more cost-effective treatment. While some medications only produce side effects or only fungistatic effects, photodynamic therapy (PDT) consists of the destruction of visible light (VL) living tissue in the presence of a photosensitizer (PS) and molecular oxygen (O₂) by response of reactive oxygen species (ROS) and singlet oxygen (¹O₂)^[17,18]. Due to the adverse effects and setbacks observed in the need for a safe therapeutic option for nail had, the study aims to investigate the action of exogenous Protoporphyrin IX (Pp IX) associated with visible light on *T. rubrum*, contemplating clinical data and providing readers with an update on current approaches for the treatment of onychomycosis by PDT.

2. Insensibility of Conventional Treatment of Onychomycosis Associated with *T. rubrum*

The standard treatment for nail had, consists of the use of local (topical) and systemic (oral) drugs or a combination of both for 4 to 6 months. Due to the long period of drug treatment, the patient ends up generating lack of interest in therapy and, thus, the patient's adherence to treatment becomes ineffective and usually promotes side effects such as liver and kidney failure^[19].

Usually, the patient seeks professional help when there are problems associated with onychomycosis, which include discomfort, difficulty wearing and wearing shoes, walking and aesthetic embarrassment in the nail segments in lower and upper fingers^[20]. Infected nails can serve as a reservoir of fungi with potential spread to the feet, hands and groin and can spread to other family members if left untreated. It is important to emphasize that onychomycosis can result in disruption of skin integrity, providing a gateway to microorganisms that lead to the development of foot ulcers (diabetic foot), osteomyelitis, cellulitis and gangrene in diabetic patients^[21]. In addition, the presence of sensitizing fungal/dermatophytic antigens in the nail lamina may predispose to other clinical conditions in individuals with onychomycosis. This includes asthma/sensitization of the respiratory tract and skin diseases such as atopic dermatitis, urticaria and erythema nodosum^[22].

2.1. Local and Oral Therapy

For mild infections, local options are highlighted, amorolfine glazes 5% and cyclopirox 8%. Amorolfina is applied once a week, while cyclopirox is applied once a day^[23]. Amorolfina has fungistatic and fungicide properties against dermatophytes, NDF and yeasts^[24], but this drug is recommended for nail without matrix involvement and mild cases of distal and lateral subungual onychomycosis (DLSO) affecting up to two nails^[25]. Cyclopyrus is a derivative of hydroxypyridon with broad-spectrum antifungal activity against *T. rubrum*, inhibiting metal-dependent enzymatic processes, including nutrient absorption, cell energy production, and intracellular peroxide degradation^[26]. However, this last drug has a lower cure rate, with side effects such as periunual erythema and nail fold^[22,27]. Therefore, the use of topical antifungals has low penetration in the nail unit, recurrences and reinfections are common^[28].

Failure of topical treatment is indicated oral therapy with terbinafine (Lamisil), itraconazole (Sporanox, Sporaz, Orungal) and fluconazole (Diflucan, Celozol). Terbinafine, an antifungal agent of the alilamine group, inhibits fungal epoxidase esqualene, leading to the accumulation of squalene, responsible for its fungicide effect^[29]. This drug is metabolized by cytochrome P450 enzymes (CYP450) and its concentration is decreased by rifampicin and increased by cimetidine^[30] and warfarin can be potentiated and may present hemorrhagic events^[31,32]. Therefore, drug interaction is limited and should be careful with the therapeutic combination, as it may affect the pharmacokinetic profile of other drugs.

This drug can be administered at 250 mg/day for 12 weeks or dosage of 500 mg/day for four weeks with pulse therapy and four weeks without pulse therapy^[33]. During treatment, common adverse reactions may appear (Table 1). Severe adverse reactions are rare, but agranulocytosis, hepatitis, acute generalized exanthematic pustulosis and lupus erytheamsus^[34] may arise.

Fluconazole is used at the dosage of 150-300 mg/week for more than six months, but is less effective^[35]. It is essential to state that this drug has not been approved by the U.S. Food and Drug Administration (FDA) for nail, although it is approved in Europe for the treatment of onychomycosis^[36,37], for representing a third-line therapy^[38]. This drug has a long half-life, is predominantly excreted in urine and remains detectable on the nails for up to 6 months after discontinuation of therapy^[39]. A high dose of 450 mg/week is observed for onychomycosis, believed to be due to its pharmacokinetic properties and advantage of improving treatment and reducing treatment costs^[22].

The common adverse effects of fluconazole (Table 1) lead to discontinuation of fluconazole therapy 150 mg weekly or higher weekly doses, such as 300-450 mg^[40]. Severe reactions to fluconazole are very rare, but occur during therapy such as polyneuropathy, thrombotic thrombocytopenic purpura, lower gastrointestinal tract bleeding, acute confusional state, and acute renal failure^[41]. This same drug presents drug interactions, such as the use of rifampicin and cimetidine, which may present possible reductions in fluconazole levels and warfarin also presents hemorrhagic events^[31,32].

Itraconazole is a triazole antifungal with a broad spectrum of activity against dermatophytes, yeasts and NDF, administered at a dosage of 400 mg/day for one week/month, with treatment duration of three months for the nails. This medicine is highly lipophilic and bioavailability increases after a meal. Seven days after the start of therapy, itraconazole is incorporated into the nail and detectable in the matrix and nail bed, and persists for up to 6 to 9 months after treatment^[30,35]. This medicine is linked to the cytochrome P450 system (CYP3A4) and is responsible for the potential toxicity of itraconazole and drug interactions. In addition to these factors, adverse reactions are manifested (Table 1). After 4 weeks of continuous therapy, hepatitis tends to occur and severe liver dysfunction can be triggered, requiring liver transplantation^[42].

As a result of this reality, monitoring of liver functions is recommended in patients who have been on continuous therapy for more than four weeks, associated with hepatotoxic drugs and pre-existing diseases. In case of congestive heart failure, itraconazole is contraindicated because of the increased risk of negative inotropic effects^[43].

Itraconazole has significant drug interactions, so it should not be taken with benzodiazepines such as midazolam (Versed[®]) and triazolam (Halcion[®]), due to overaire sedation^[31,32].

Compared to another drug, itraconazole is a stronger inhibitor of cytochrome P450 enzymes than fluconazole^[44], however, both have more side effects and potential drug interactions than terbinafine^[7,45,46].

Table 1. Adverse effects of systemic drugs during onychomycosis therapy.

Drugs	Drug whose metabolism is affected	Reactions	
		Common/Mild	Severe/Rare
Terbinafine	Cimetidine, rifampicin and warfarin	Rash, itching, hives, gastrointestinal symptoms, taste and liver enzyme changes.	Agranulocytosis, hepatitis, acute generalized exanthematous pustulosis, and lupus erythematosus
Fluconazole	Rifampicin, cimetidine and warfarin	Headache, rash, gastrointestinal complaints, insomnia, paraesthesia and dysesthesia	PNP, lower gastrointestinal bleeding, PTT, ARF and ACE
Itraconazole	Midazolam and triazolam	Headache and gastrointestinal symptoms	Hepatitis and severe liver dysfunction

Legend: polyneuropathy (PNP); thrombotic thrombocytopenic purpura (PTT); acute renal failure (ARF); acute confusional state (ACE).

2.2. Cognitive Considerations in Relation to Therapy

In severe cases of onychomycosis, such as LNP involvement, dermatophytomas or dystrophic onychomycosis, surgical or chemical intervention of the nail lamina associated with topical or systemic treatment with itraconazole and terbinafine^[47] is indicated.

Treatment of onychomycosis requires several months of therapy, since nail growth is very slow, especially in the elderly. The choice of medication depends on the type and severity of onychomycosis and the comorbidities of the patient. In most cases, patients have DLSO due to dermatophytes involving the distal part of one or two large nails^[48].

However, the use of systemic drugs is limited by hepatotoxicity and drug interactions, representing a safety concern, especially in people over 60 years of age. A 20% failure rate of oral antifungals and a high recurrence rate of 10-53 stand out. Factors that also contribute to the failure of therapy are patient susceptibility, resistant fungal growth pattern, presence of latent fungal spores on the nail, low bioavailability of the drug and lack of penetration of the drug into the nail^[22,49-51].

The insensitivity of nail infection to antifungal therapy often leads to the need for an alternative therapy to fight *T. rubrum*, so researchers, as well as dermatologists, seek a beneficial treatment for nail had, a fruitful therapy with toxic levels only for the microbial agent, minimal side effects and low cost for the patient so that he does not give up therapy during the course of treatment, because of this, PDT is a therapy capable of consolidating the therapeutic option for the control of the fungal agent.

3. Drug-Induced Liver Injury in Nail Infection

The liver is a site where several medications are metabolized, a classic example are antifungals. One type of antifungal that deserves to be highlighted is terbinafine. This medicine is used to combat fungi causing onychomycosis, but liver injury may occur during treatment, damage known as drug-induced liver injury (DILI)^[52,53]. DILI is defined as liver damage induced by a commercial or herbal medicine that leads to liver dysfunction^[54].

This factor denotes that oral antifungals are associated with an incidence of DILI, about 4% for terbinafine, 1% for fluconazole and 2 to 4% for itraconazole^[55]; however, these drugs can be fatal, especially in the elderly and long-term treatments^[56].

The diagnosis of liver injury is a major challenge for physicians and is based on a combination of factors such as exclusion from other causes that can elevate biochemical liver tests and causality assessment methods, which can be based on expert opinion or standardized methods specific to the liver. In this sense, the Roussel Uclaf Causality Assessment Method (RUCAM) is indicated^[53,57]. Currently, RUCAM is an instrument used to assist in determining specific causality of DILI, thus specifically assessing the probability of high levels of drug-induced liver-associated enzymes^[52,58].

4. Photodynamic Therapy

Because it is a broad and complex field of a multidisciplinary nature (biochemistry, materials science, chemistry, engineering and medicine), PDT has been investigated for more than 100 years and, for this reason, scientific research has grown exponentially in the last two decades. PDT was introduced to act as a complementary approach to conventional therapies to reduce the recurrence of cellular damage and extend survival with minimal side effects^[59-61]. Thus, it can be considered that PDT is a consolidated therapeutic modality that has its activity in the photooxidation (destruction of target cells) of biological matter, which results from the activity of ROS and ¹O₂ (Figure 1). These species are generated in situ and cause cell death by LV (Figure 2) in the presence of PS and O₂^[62].

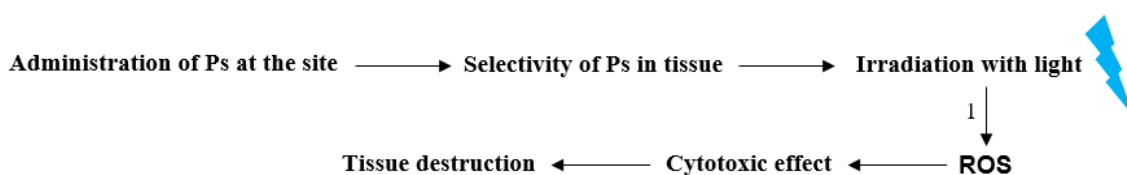


Figure 1. PDT simplified flowchart. Caption: (1) Photodynamic action; (ROS) reactive oxygen species.

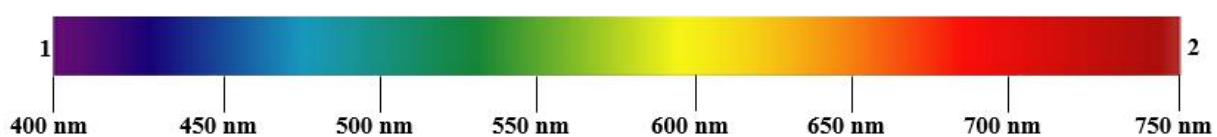


Figure 2. Visible spectrum of light (400-750 nm). **Legend:** (nm) nanometer; (1) ultraviolet spectrum below 400 nm; (2) infrared spectrum above 750 nm.

4.1. Photosensitizer “Protoporphyrin IX”

Currently, there are several PS for PDT, but few studies on basic structure and its activity, which makes the investigation of structure and activity of great interest in terms of photobiology. It is also added that the clinical future for the application of PDT will be highly dependent on efficient PS^[63,64] with minimal toxicity in the dark, low mutagenic potential and highly chemically stable^[65,66], which will contribute to the advancement and popularization of PDT. Regarding the previous approach, there is a need to seek PS of commercial origin and/or derived from low-cost synthesis, which provides better permeability and incorporation in both tissues and cells, with reduced photosensitive reaction capable of curing the lesion with minimal adverse reactions and restoring the normal structure and function of the tissue.

PS "Pp IX" (Figure 3) is a dye with the ability to absorb energy from one light source and transfer this energy to another molecule, presenting photodynamic activity^[67,68]. The solubility of this PS appears to be one of the characteristics of relevance, since the photochemical and pharmacokinetic properties depend on it. Another relevant factor is that it occupies a special place in the porphyrin family due to its important role in various clinical-therapeutic applications, including PDT^[69,70] for adhering to membranes and cellular components^[71,72] and for having an intense light absorption peak around 405 nm (Soret band) and a much weaker absorption band at 635 nm^[73], which allows its activation *in vivo* by red light, while blue, green or yellow light are used in superficial applications, for example on the skin^[74-76], which makes Pp IX viable for both dermatological treatment and antimicrobial photodynamic inactivation.

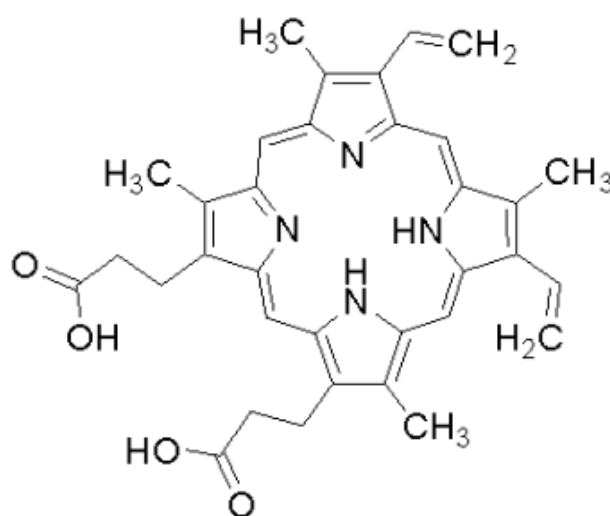


Figure 3: Molecular structure of Pp IX.

4.2. Mechanism of Action

The photodynamic process is initiated by the absorption of radiant energy. This irradiation should occur at an adequate wavelength (λ), i.e., PS should be irradiated in a spectral region where it has absorption bands. By absorbing a photon, PS reaches the excited state and, later, the chain of dynamic processes described in the modified Jablonski diagram occurs (Figure 4). In this process, different light sources can be used, such as white sunlight from solar irradiation (natural light) or preferably an artificial irradiation source that emits in the visible region (light-emitting diode (LED) or Lasers). However, the photodynamic process will be

proportional to the number of photons absorbed by the PS in the appropriate λ and consequently proportional to the intensity of emission of the source in the absorption λ of the respective PS^[75,77,78].

In the photodynamic process, the PS that is in the fundamental state absorbs radiant energy by moving into the singlet excited state ($^1PS^*$). After excitation, PS is driven to the excited triplet state ($^3PS^*$) or can return to the fundamental state by phosphorescence without chemical alteration. In $^3PS^*$ the species can generate ROS. These species can be generated by abstraction or absorption of electrons or hydrogens, leading to the formation of radicals (mechanism of action of type I). Also by deactivating $^3PS^*$ by transferring energy to 3O_2 leading to the formation of 1O_2 (type II mechanism of action), both mechanisms (Figure 4) promote cell death^[79-82].

PS and λ of the irradiation source are two inseparable elements in the therapeutic process of PDT, however, the selection of these elements are also related to the depth of tissue to be treated. In treatment with PDT, ultraviolet (UV) and infrared (IR) irradiation (IR) is excluded; UV for causing tissue damage and IR for reaching heat waves, being restricted to the LV region. For treatments to be performed in regions of greater depth in the tissue, it is necessary to radiate in red or near infrared, where light reaches greater penetration into the biological tissue. On the other hand, the treatments to be performed on the epidermis or dermis do not require λ greater penetration into the tissue, in this case, the use of blue and violet light is indicated for therapy, for not harming deeper tissues^[83-85].

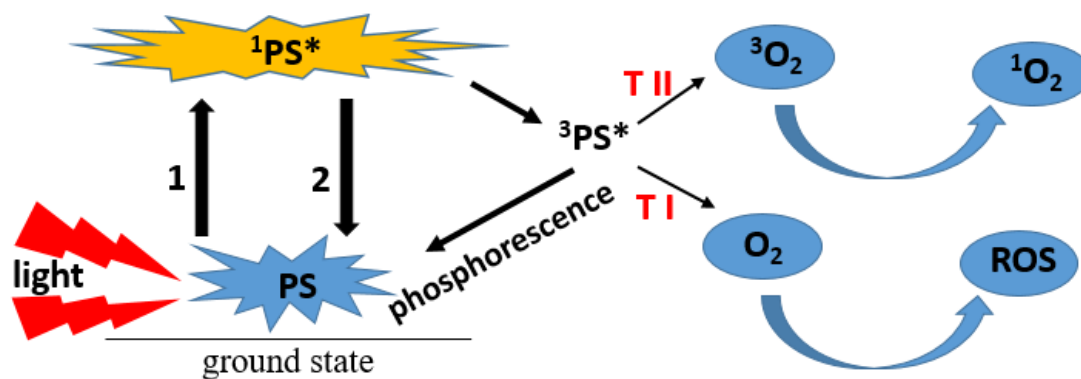


Figure 4: Simplified Jablonski diagram. Mechanism Type I and Type II of photosensitization. Caption: (PS) ground state photosensitizer; (1) absorption of radiant energy; ($^1PS^*$) singlet excited state; (2) excited photosensitizer can return to ground state as fluorescence; ($^3PS^*$) triplet state; (T I) Type I Mechanism; (T II) Type II Mechanism; (3O_2) triplet oxygen; (1O_2) singlet oxygen; (ROS) reactive oxygen species.

5. Photodynamic Action of the Photosensitizer Associated with Visible Light

Ramos *et al.*^[18] demonstrated in their study the Pp IX dimethyl ester obtained by derivatization of Pp IX in an analogous procedure described by Uchoa *et al.*^[86]. In the *in vitro* study by Ramos *et al.*, Pp IX homogenized with *T. rubrum* was irradiated with blue LED ≈ 400 nm. The samples were submitted to 12 cycles of 10 minutes of irradiation with LED. The authors reported that the control group had an average growth of more than 3.000 colony-forming units (CFU). On the other hand, the sample of *T. rubrum* + Pp IX with irradiation, there was an intensified reduction in the first six cycles. No significant differences were observed between the seventh and the other subsequent cycles, remaining between 0 and 1 CFU. Another

aspect, also very important, is that the authors described the incorporation of PS into *T. rubrum* by fluorescence microscopy. The internalization of Pp IX in the cell resulted in photooxidative damage when irradiated with λ of 400 nm, region in which this PS presents the band with the highest absorption intensity, the Soret band. The incorporation and internalization of PS in *T. rubrum* shows that the elimination of the fungal agent occurred by a photoinduced process, since this process is initiated by the transfer of hydrogens or electrons, radical reactions or electrophilic attack of singlet oxygen in cell membranes^[18,87].

An advantage of the use of blue light (400-480 nm) is that this λ can reduce the proliferative activity of keratinocytes, modulating the immune responses of T cells and safely improving superficial infections^[88,89].

Another approach to reduce the infectious process is red light with λ from 620 to 770 nm for deeper treatment of up to 6 mm, being able to stimulate mitochondrial activity and modulate the release of cytokines from macrophages^[90,91].

The nanomaterial called carbon points (CDs) stands out. CDs are a new class of carbon-based fluorescent nanoparticles with physical and chemical characteristics. Researchers like Wu *et al.*^[92] and Kang *et al.*^[93] consider CDs an ideal candidate for drug delivery applications due to their photophysical properties, low toxicity, adjustable surface functionality and adaptive synthesis^[92,93]. These particularities of CDs together with PS have gained research interest in PDT^[94].

Amide crosslinking is a strategy that is based on interactions between carboxyl and primary amine groups. The factor that favors this bond is the EDC (1-ethyl-3-(3-dimethylamino) carbodiimide propyl). EDC is a zero-length reticulative that can be used in conjunction with N-hydroxyuccinimide (NHS) to efficiently produce amide bonds between two suitable molecules^[95,96]. An *in vitro* study shows that Pp IX was previously covalently linked through carbodiimide chemistry, and also excitation of two photons and singlet oxygen, which evidences an important mechanism for PDT^[97-101].

The incipience of scientific articles published on the proposed theme is in poor development not only in dermatology, but also in medical clinic and other areas of health, which is why it is essential to develop and publish exogenous Pp IX studies on *T. rubrum* with visible spectrum light treatment, to propagate current knowledge about a specific therapy of PDT for onychomycosis.

Table 2 summarizes the final results of the therapy, but the PS acid 5-aminolevulinic acid (ALA). Endogenous photosensitizer, Pp IX, produced from its precursor ALA in the heme biosynthesis pathway is also important in the case of antimicrobial PDT application^[102,103].

Table 2. Summary of the various forms of dermatophytosis treated with PDT.

Disease	Fungal species	PS	Final result	Reference
Interdigital mycoses	<i>T. mentagrophytes</i> <i>T. rubrum</i>	ALA (20% cream solution)	No clinical signs	Calzavara-Pinton <i>et al.</i> ^[104]
UR	<i>T. rubrum</i>	ALA	Growth inhibitory effect	Kamp <i>et al.</i> ^[105]
Onychomycosis	<i>T. rubrum</i>	ALA	Clinical and mycological healing	Piraccini <i>et al.</i> ^[106]
<i>Tinea cruris</i>	<i>T. rubrum</i>	ALA	100% healing rate	Sotiriou <i>et al.</i> ^[107]

Foot Tinea	<i>T. mentagrophytes</i> var.interdigitales <i>T. rubrum</i>	ALA cream	Absence of mycological signs after 3 treatments	Sotiriou <i>et al.</i> ^[108]
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Legend: 5-aminolevulinic acid (ALA); unrevealed (UR).

6. Clinical and Future Perspectives

Light-on-tissue treatment is a safe, easy and effective therapeutic modality for various dermatoses, with lower cost, minimal side effects and superior efficacy compared to local or oral therapy in the treatment of patients with mycosis, atopic dermatitis, fungicides and others such as psoriasis, pityriasis, versicolor, chronic urticaria, palmoplantar pustulose and vitiligo. This therapeutic option prevents patients from suffering the side effects of local/oral therapy because it is a safe alternative without microbial resistance to the patient^[109,110].

In addition, PDT has great potential in the field of tumor treatment due to its advantages, such as high treatment efficiency and strong selectivity, and brings new favorable clinical aspects, as well as a good prognosis and opportunities for future tumor therapies^[111]. Another significant point is that PDT can activate the immune system through molecular patterns associated with damage (DAMPs) released or exposed to dead cells, which in turn stimulate innate and adaptive immunity, this being the first line of defense of the organism^[61,112-114].

There is not much *in vitro* and *in vivo* research published on the photodynamic process of exogenous Pp IX with the combination of light devices in the visible spectrum for the inactivation of *T. rubrum*. Future issues, such as the high success rate and short-term therapy associated with Pp IX and light, should be researched to develop a safe protocol for the treatment of *T. rubrum* causing onychomycosis.

7. Conclusion

Drug treatment of onychomycosis requires several months of therapy, in addition to presenting adverse effects and recurrences. Due to these factors, PDT is a recommended therapeutic alternative for onychomycosis, because it does not present microbial resistance with minimal adverse effects at the site. Pp IX together with visible light showed antimicrobial activity against *T. rubrum in vitro*, so it is suggested that PS may be a promising candidate in PDT for the treatment of onychomycosis. But since there is little evidence of Pp IX photosensitization on *T. rubrum*, more research with significant results is needed to justify the photodynamic process. Given this gap, research with elements of PDT (Pp IX, visible light and molecular oxygen "*T. rubrum*") are essential to develop and offer a therapy that not only satisfies the desire of the patient with onychomycosis, but also surprises him with healing and aesthetics, exceeding his expectations and improving the quality of life.

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DECLARATION OF POTENTIAL CONFLICT OF INTEREST

The authors declare no conflict of interest.

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References

- [1] Ghannoum M, Isham N. Fungal nail infections (onychomycosis): a never-ending story? PLoS Pathog 2014; 10(6): e1004105. <https://doi.org/10.1371/journal.ppat.1004105>
- [2] Gupta AK, Versteeg SG, Shear NH. Onychomycosis in the 21st century: an update on diagnosis, epidemiology, and treatment. J Cutan Med Surg 2017; 21(6): 525-539. <https://doi.org/10.1177/1203475417716362>
- [3] Poulakos M, Grace Y, Machin JD, Dorval E. Efinaconazole and tavaborole. J Pharm Pract 2017; 30(2): 245-255. <https://doi.org/10.1177/0897190016630904>
- [4] Pakshir K, Kamali M, Nouraei H, Zomorodian K, Motamedi M, Mahmoodi M. Molecular characterization and antifungal activity against non-dermatophyte molds causing onychomycosis. Sci Rep 2021;11(1): 20736. <https://doi.org/10.1038/s41598-021-00104-0>
- [5] Lipner SR, Scher RK. Onychomycosis: clinical overview and diagnosis. J Am Acad Dermatol 2019; 80(4): 835-851. <https://doi.org/10.1016/j.jaad.2018.03.062>
- [6] Grover C, Khurana A. Onychomycosis: newer insights in pathogenesis and diagnosis. Indian J Dermatol Venereol Leprol 2012; 78(3): 263-270. <https://doi.org/10.4103/0378-6323.95440>
- [7] Leung AKC, Lam JM, Leong KF, *et al.* Onychomycosis: an updated review. Recent Pat Inflamm Allergy Drug Discov 2020; 14(1): 32-45. <https://doi.org/10.2174/1872213X13666191026090713>
- [8] Peres NT, Maranhão FC, Rossi A, Martinez-Rossi NM. Dermatophytes: host-pathogen interaction and antifungal resistance. An Bras Dermatol 2010; 85: 657-667. <https://doi.org/10.1590/s0365-05962010000500009>
- [9] Nir-Paz R, Elinav H, Pierard GE, *et al.* Deep infection by *Trichophyton rubrum* in an immunocompromised patient. J Clin Microbiol 2003; 41: 5298-52301. <https://doi.org/10.1128/JCM.41.11.5298-5301.2003>
- [10] Maranhão FC, Paião FG, Fachin AL, Martinez-Rossi NM. Membrane transporter proteins are involved in *Trichophyton rubrum* pathogenesis. J Med Microbiol 2009; 58: 163-168. <https://doi.org/10.1099/jmm.0.002907-0>
- [11] Giddey K, Monod M, Barblan J, *et al.* Comprehensive analysis of proteins secreted by *Trichophyton*

- rubrum* and *Trichophyton violaceum* under *in vitro* conditions. J Proteome Res, 2007; 6(8): 3081-3092. <https://doi.org/10.1021/pr070153m>
- [12] Sriranganadane D, Waridel P, Salamin K, *et al.* Identification of novel secreted proteases during extracellular proteolysis by dermatophytes at acidic pH. Proteomics 2011; 11(22): 4422-4433. <https://doi.org/10.1002/pmic.201100234>
- [13] Monod M, Méhul B. Recent findings in onychomycosis and their application for appropriate treatment. J Fungi (Basel) 2019; 5(1): 20. <https://doi.org/10.3390/jof5010020>
- [14] Dharmoon RK, Popli H, Gupta M. Novel drug delivery strategies for the treatment of onychomycosis. Pharm Nanotechnol 2019; 7(1): 24-38. <https://doi.org/10.2174/2211738507666190228104031>
- [15] Gupta AK, Foley KA. Evidence for biofilms in onychomycosis. G Ital Dermatol Venereol 2019; 154(1): 50-55. <https://doi.org/10.23736/S0392-0488.18.06001-7>
- [16] Girois SB, Chapuis F, Decullier E, Revol BG. Adverse effects of antifungal therapies in invasive fungal infections: review and meta-analysis. Eur J Clin Microbiol Infect Dis 2006; 25: 138-149. <https://doi.org/10.1007/s10096-005-0080-0>
- [17] Rodrigues GB, Ferreira LK, Wainwright M, Braga GU. Susceptibilities of the dermatophytes *Trichophyton mentagrophytes* and *T. rubrum* microconidia to photodynamic antimicrobial chemotherapy with novel phenothiazinium photosensitizers and red light. J Photochem Photobiol B 2012; 116: 89-94. <https://doi.org/10.1016/j.jphotobiol.2012.08.010>
- [18] Ramos RR, Kozusny-Andreani DI, Fernandes AU, Baptista MS. Photodynamic action of protoporphyrin IX derivatives on *Trichophyton rubrum*. An Bras Dermatol 2016; 91(2): 135-140. <https://doi.org/10.1590/abd1806-4841.20163643>
- [19] Morgado LF, Trávolo ARF, Muehlmann LA, *et al.* Photodynamic therapy treatment of onychomycosis with aluminium-phthalocyanine chloride nanoemulsions: a proof of concept clinical trial. J Photochem Photobiol B 2017; 173: 266-270. <https://doi.org/10.1016/j.jphotobiol.2017.06.010>
- [20] Thomas J, Jacobson GA, Narkowicz CK, Peterson GM, Burnet H, Sharpe C. Toenail onychomycosis: an important global disease burden. J Clin Pharm Ther 2010; 35(5): 497-519. <https://doi.org/10.1111/j.1365-2710.2009.01107.x>
- [21] Tan JS, Joseph WS. Common fungal infections of the feet in patients with diabetes mellitus. Drugs Aging 2004; 21(2): 101-112. <https://doi.org/10.2165/00002512-200421020-00003>
- [22] Ameen M, Lear JT, Madan V, Mohd Mustapa MF, Richardson M. British Association of Dermatologists' guidelines for the management of onychomycosis 2014. Br J Dermatol 2014; 171(5): 937-958. <https://doi.org/10.1111/bjd.13358>
- [23] Gupta AK, Daigle D, Foley KA. Topical therapy for toenail onychomycosis: an evidence-based review. Am J Clin Dermatol 2014; 15(6): 489-502. <https://doi.org/10.1007/s40257-014-0096-2>
- [24] Gupta AK, Ryder JE, Baran R. The use of topical therapies to treat onychomycosis. Dermatol Clin 2003; 21(3): 481-489. [https://doi.org/10.1016/s0733-8635\(03\)00025-1](https://doi.org/10.1016/s0733-8635(03)00025-1)
- [25] Gupta AK, Paquet M, Simpson FC. Therapies for the treatment of onychomycosis. Clin Dermatol 2013; 31(5): 544-554. <https://doi.org/10.1016/j.clindermatol.2013.06.011>
- [26] Bohn M, Kraemer KT. Dermatopharmacology of ciclopirox nail lacquer topical solution 8% in the

- treatment of onychomycosis. *J Am Acad Dermatol* 2000; 43(4 Suppl): S57-69. <https://doi.org/10.1067/mjd.2000.109072>
- [27] Shemer A, Nathansohn N, Trau H, Amichai B, Grunwald MH. Ciclopirox nail lacquer for the treatment of onychomycosis: an open non-comparative study. *J Dermatol* 2010; 37(2): 137-139. <https://doi.org/10.1111/j.1346-8138.2009.00773.x>
- [28] Del Rosso JQ. The role of topical antifungal therapy for onychomycosis and the emergence of newer agents. *J Clin Aesthet Dermatol* 2014; 7(7): 10-18. Available from: <https://pubmed.ncbi.nlm.nih.gov/25053979/>
- [29] Sigurgeirsson B, Billstein S, Rantanen T, *et al.* L.I.ON. Study: efficacy and tolerability of continuous terbinafine (Lamisil) compared to intermittent itraconazole in the treatment of toenail onychomycosis. Lamisil vs. itraconazole in onychomycosis. *Br J Dermatol* 1999; 141(Suppl 56): 5-14. <https://doi.org/10.1046/j.1365-2133.1999.00008.x>
- [30] Singal A, Khanna D. Onychomycosis: diagnosis and management. *Indian J Dermatol Venereol Leprol* 2011; 77(6): 659-672. <https://doi.org/10.4103/0378-6323.86475>
- [31] Katz HI, Gupta AK. Oral antifungal drug interactions. *Dermatol Clin* 1997; 15(3): 535-544. [https://doi.org/10.1016/s0733-8635\(05\)70460-5](https://doi.org/10.1016/s0733-8635(05)70460-5)
- [32] Gupta AK, Versteeg SG, Shear NH. Common drug-drug interactions in antifungal treatments for superficial fungal infections. *Expert Opin Drug Metab Toxicol* 2018; 14(4): 387-398. <https://doi.org/10.1080/17425255.2018.1461834>
- [33] Gupta AK, Paquet M, Simpson F, Tavakkol A. Terbinafine in the treatment of dermatophyte toenail onychomycosis: a meta-analysis of efficacy for continuous and intermittent regimens. *J Eur Acad Dermatol Venereol* 2013; 27(3): 267-272. <https://doi.org/10.1111/j.1468-3083.2012.04584.x>
- [34] Bonsmann G, Schiller M, Luger TA, Ständer S. Terbinafine-induced subacute cutaneous lupus erythematosus. *J Am Acad Dermatol* 2001; 44(6): 925-931. <https://doi.org/10.1067/mjd.2001.114565>
- [35] Gupta AK, Drummond-Main C, Paquet M. Evidence-based optimal fluconazole dosing regimen for onychomycosis treatment. *J Dermatolog Treat* 2013; 24(1): 75-80. <https://doi.org/10.3109/09546634.2012.703308>
- [36] Lipner SR, Scher RK. Onychomycosis: clinical overview and diagnosis. *J Am Acad Dermatol* 2019; 80(4): 835-851. <https://doi.org/10.1016/j.jaad.2018.03.062>
- [37] Lipner SR, Scher RK. Onychomycosis: treatment and prevention of recurrence. *J Am Acad Dermatol* 2019; 80(4): 853-867. <https://doi.org/10.1016/j.jaad.2018.05.1260>
- [38] Rendl M, Mayer C, Weninger W, Tschachler E. Topically applied lactic acid increases spontaneous secretion of vascular endothelial growth factor by human reconstructed epidermis. *Br J Dermatol* 2001; 145(1): 3-9. <https://doi.org/10.1046/j.1365-2133.2001.04274.x>
- [39] Rich P, Scher RK, Breneman D, *et al.* Pharmacokinetics of three doses of once-weekly fluconazole (150, 300, and 450 mg) in distal subungual onychomycosis of the toenail. *J Am Acad Dermatol* 1998; 38(6 Pt 2): S103-109. [https://doi.org/10.1016/s0190-9622\(98\)70493-1](https://doi.org/10.1016/s0190-9622(98)70493-1)
- [40] Chang CH, Young-Xu Y, Kurth T, Orav JE, Chan AK. The safety of oral antifungal treatments for superficial dermatophytosis and onychomycosis: a meta-analysis. *Am J Med* 2007; 120(9): 791-798. <https://doi.org/10.1016/j.amjmed.2007.03.021>

- [41] Eşkut N, Gedizlioğlu M, Ünal O, Özlü C, Ergene U. Acute fluconazole toxicity: a case presenting with protean manifestations including systemic and neurologic symptoms. *Postgrad Med* 2021; 133(2): 250-252. <https://doi.org/10.1080/00325481.2020.1840830>
- [42] Srebrnik A, Levtov S, Ben-Ami R, Brenner S. Liver failure and transplantation after itraconazole treatment for toenail onychomycosis. *J Eur Acad Dermatol Venereol* 2005; 19(2): 205-207. <https://doi.org/10.1111/j.1468-3083.2005.00943.x>
- [43] Cathcart S, Cantrell W, Elewski Be. Onychomycosis and diabetes. *J Eur Acad Dermatol Venereol* 2009; 23(10): 1119-1122. <https://doi.org/10.1111/j.1468-3083.2009.03225.x>
- [44] Niwa T, Shiraga T, Takagi A. Effect of antifungal drugs on cytochrome P450 (CYP) 2C9, CYP2C19, and CYP3A4 activities in human liver microsomes. *Biol Pharm Bull* 2005; 28(9): 1805-1808. <https://doi.org/10.1248/bpb.28.1805>
- [45] Hoy NY, Leung AK, Metelitsa AI, Adams S. New concepts in median nail dystrophy, onychomycosis, and hand, foot, and mouth disease nail pathology. *ISRN Dermatol* 2012; 2012: 680163. <https://doi.org/10.5402/2012/680163>
- [46] Bodman MA, Krishnamurthy K. Onychomycosis. In: *StatPearls*. Treasure Island (FL): StatPearls Publishing; 2021. Available from: <https://pubmed.ncbi.nlm.nih.gov/28722883/>
- [47] Lecha M, Effendy I, Feuilhade de Chauvin M, Di Chiacchio N, Baran R; Taskforce on onychomycosis education. Treatment options--development of consensus guidelines. *J Eur Acad Dermatol Venereol* 2005; 19(Suppl 1): 25-33. <https://doi.org/10.1111/j.1468-3083.2005.01284.x>
- [48] Piraccini BM, Alessandrini A. Onychomycosis: a review. *J Fungi (Basel)* 2015; 1(1): 30-43. <https://doi.org/10.3390/jof1010030>
- [49] Evans EG. The rationale for combination therapy. *Br J Dermatol* 2001; 145(Suppl 60): 9-13. <https://pubmed.ncbi.nlm.nih.gov/11777263/>
- [50] Tabara K, Szewczyk AE, Bienias W, *et al.* Amorolfine vs. ciclopirox - lacquers for the treatment of onychomycosis. *Postepy Dermatol Alergol* 2015; 32(1): 40-45. <https://doi.org/10.5114/pdia.2014.40968>
- [51] Olafsson JH, Sigurgeirsson B, Baran R. Combination therapy for onychomycosis. *Br J Dermatol* 2003; 149(Suppl 65): 15-18. <https://doi.org/10.1046/j.1365-2133.149.s65.2.x>
- [52] Paredes AH, Lewis JH. Terbinafine-induced acute autoimmune hepatitis in the setting of hepatitis B virus infection. *Ann Pharmacother* 2007; 41(5): 880-884. <https://doi.org/10.1345/aph.1H400>
- [53] Raschi E, Poluzzi E, Koci A, Caraceni P, Ponti FD. Assessing liver injury associated with antimycotics: concise literature review and clues from data mining of the FAERS database. *World J Hepatol* 2014; 6(8): 601-612. <https://doi.org/10.4254/wjh.v6.i8.601>
- [54] Hussaini SH, Farrington EA. Idiosyncratic drug-induced liver injury: an update on the 2007 overview. *Expert Opin Drug Saf* 2014; 13(1): 67-81. <https://doi.org/10.1517/14740338.2013.828032>
- [55] Song JC, Deresinski S. Hepatotoxicity of antifungal agents. *Curr Opin Investig Drugs* 2005; 6(2): 170-177. Available from: <https://pubmed.ncbi.nlm.nih.gov/15751740/>
- [56] Kao WY, Su CW, Huang YS, *et al.* Risk of oral antifungal agent-induced liver injury in Taiwanese. *Br J Clin Pharmacol* 2014; 77(1): 180-189. <https://doi.org/10.1111/bcp.12178>
- [57] Teschke R, Wolff A, Frenzel C, Schwarzenboeck A, Schulze J, Eickhoff A. Drug and herb induced

- liver injury: council for International Organizations of Medical Sciences scale for causality assessment. *World J Hepatol* 2014; 6(1): 17-32. <https://doi.org/10.4254/wjh.v6.i1.17>
- [58] LiverTox: Clinical and Research Information on Drug-Induced Liver Injury. Bethesda (MD): National Institute of Diabetes and Digestive and Kidney Diseases; Roussel Uclaf Causality Assessment Method (RUCAM) in Drug Induced Liver Injury. *LiverTox* 2012; 1-8. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK548272/>
- [59] Bechet D, Mordon SR, Guillemin F, Barberi-Heyob MA. Photodynamic therapy of malignant brain tumours: a complementary approach to conventional therapies. *Cancer Treat Rev* 2014; 40(2): 229-241. <https://doi.org/10.1016/j.ctrv.2012.07.004>
- [60] Akimoto J. Photodynamic therapy for malignant brain tumors. *Neurol Med Chir* 2016; 56(4): 151-157. <https://doi.org/10.2176/nmc.ra.2015-0296>
- [61] Algorri JF, Ochoa M, Roldán-Varona P, Rodríguez-Cobo L, López-Higuera JM. Photodynamic therapy: a compendium of latest reviews. *Cancers (Basel)* 2021; 13(17): 4447. <https://doi.org/10.3390/cancers13174447>
- [62] Inada NM, Costa MM, Guimarães OC, *et al.* Photodiagnosis and treatment of condyloma acuminatum using 5-aminolevulinic acid and homemade devices. *Photodiagnosis Photodyn Ther* 2012; 9(1): 60-68. <https://doi.org/10.1016/j.pdpdt.2011.09.001>
- [63] Savellano MD, Hasan T. Targeting cells that overexpress the epidermal growth factor receptor with polyethylene glycolated BPD verteporfin photosensitizer immunoconjugates. *Photochem Photobiol* 2003; 77(4): 431-439. [https://doi.org/10.1562/0031-8655\(2003\)077<0431:tctote>2.0.co;2](https://doi.org/10.1562/0031-8655(2003)077<0431:tctote>2.0.co;2)
- [64] Lin Y, Zhou T, Bai R, Xie Y. Chemical approaches for the enhancement of porphyrin skeleton-based photodynamic therapy. *J Enzyme Inhib Med Chem* 2020; 35(1): 1080-1099. <https://doi.org/10.1080/14756366.2020.1755669>
- [65] Nyman ES, Hynninen PH. Research advances in the use of tetrapyrrolic photosensitizers for photodynamic therapy. *J Photochem Photobiol B* 2004; 73(1-2): 1-28. <https://doi.org/10.1016/j.jphotobiol.2003.10.002>
- [66] Plaetzer K, Krammer B, Berlanda J, Berr F, Kiesslich T. Photophysics and photochemistry of photodynamic therapy: fundamental aspects. *Lasers Med Sci* 2009; 24(2): 259-268. <https://doi.org/10.1007/s10103-008-0539-1>
- [67] Bigelow CE, Mitra S, Knuechel R, Foster TH. ALA- and ALA-hexylester-induced protoporphyrin IX fluorescence and distribution in multicell tumour spheroids. *Br J Cancer* 2001; 85(5): 727-734. <https://doi.org/10.1054/bjoc.2001.1977>
- [68] Gronlund-Pakkanen S, Wahlfors J, Makinen K, *et al.* The fluorescence biodistribution and kinetics of aminolevulinic acid induced protoporphyrin IX in the bladder of a rat model with orthotopic urothelial carcinoma. *J Urol* 2002; 167(4): 1848-1853 Available from: <https://pubmed.ncbi.nlm.nih.gov/11912446/>
- [69] Theodossiou T, MacRobert AJ. Comparison of the photodynamic effect of exogenous photoporphyrin and protoporphyrin IX on PAM 212 murine keratinocytes. *Photochem Photobiol* 2002; 76(5): 530-537. [https://doi.org/10.1562/0031-8655\(2002\)076<0530:cotpeo>2.0.co;2](https://doi.org/10.1562/0031-8655(2002)076<0530:cotpeo>2.0.co;2)

- [70] Bonnett R, Martínez G. Photobleaching of compounds of the 5,10,15,20-Tetrakis(m-hydroxyphenyl)porphyrin series (m-THPP, m-THPC, and m-THPBC). *Org Lett* 2002; 4(12): 2013-2016. <https://doi.org/10.1021/ol025842c>
- [71] Cernay T, Zimmermann HW. Selective photosensitization of mitochondria by the lipophilic cationic porphyrin POR10. *J Photochem Photobiol B* 1996; 34(2-3): 191-196. [https://doi.org/10.1016/1011-1344\(95\)07267-5](https://doi.org/10.1016/1011-1344(95)07267-5)
- [72] Zimmermann A, Ritsch-Marte M, Kostron H. mTHPC-mediated photodynamic diagnosis of malignant brain tumors. *Photochem Photobiol* 2001; 74(4): 611-616. [https://doi.org/10.1562/0031-8655\(2001\)074<0611:MMPDOM>2.0.CO;2](https://doi.org/10.1562/0031-8655(2001)074<0611:MMPDOM>2.0.CO;2)
- [73] Rollakanti KR, Kanick SC, Davis SC, Pogue BW, Maytin EV. Techniques for fluorescence detection of protoporphyrin IX in skin cancers associated with photodynamic therapy. *Photonics Lasers Med* 2013; 2(4): 287-303. <https://doi.org/10.1515/plm-2013-0030>
- [74] Calzavara-Pinton P, Rossi MT, Sala R, Venturini M. Photodynamic antifungal chemotherapy. *Photochem Photobiol* 2012; 88(3): 512-522. <https://doi.org/10.1111/j.1751-1097.2012.01107.x>
- [75] Abrahamse H, Hamblin MR. New photosensitizers for photodynamic therapy. *Biochem J* 2016; 473(4): 347-364. <https://doi.org/10.1042/BJ20150942>
- [76] Tampa M, Sarbu MI, Matei C, *et al.* Photodynamic therapy: a hot topic in dermato-oncology. *Oncol Lett* 2019; 17(5): 4085-4093. <https://doi.org/10.3892/ol.2019.9939>
- [77] Konan YN, Gurny R, Allémann E. State of the art in the delivery of photosensitizers for photodynamic therapy. *J Photochem Photobiol B* 2002; 66(2): 89-106. [https://doi.org/10.1016/s1011-1344\(01\)00267-6](https://doi.org/10.1016/s1011-1344(01)00267-6)
- [78] Rajesh S, Koshi E, Philip K, Mohan A. Antimicrobial photodynamic therapy: an overview. *J Indian Soc Periodontol* 2011; 15(4): 323-327. <https://doi.org/10.4103/0972-124X.92563>
- [79] De Rosa FS, Bentley MV. Photodynamic therapy of skin cancers: sensitizers, clinical studies and future directives. *Pharm Res* 2000; 17(12): 1447-1455. <https://doi.org/10.1023/a:1007612905378>
- [80] Donnelly RF, McCarron PA, Tunney MM. Antifungal photodynamic therapy. *Microbiol Res* 2008; 163(1): 1-12. <https://doi.org/10.1016/j.micres.2007.08.001>
- [81] Ramos RR, Paiva JL, Gomes JPFDS, Boer NP, Godoy JMP, Batigalia F. Photodynamic action of the red laser on *Propionibacterium acnes*. *An Bras Dermatol* 2017; 92(5): 622-625. <https://doi.org/10.1590/abd1806-4841.20175651>
- [82] Kwiatkowski S, Knap B, Przystupski D, *et al.* Photodynamic therapy - mechanisms, photosensitizers and combinations. *Biomed Pharmacother* 2018; 106: 1098-1107. <https://doi.org/10.1016/j.biopha.2018.07.049>
- [83] Kalka K, Merk H, Mukhtar H. Photodynamic therapy in dermatology. *J Am Acad Dermatol* 2000; 42(3): 389-413; 414-416. [https://doi.org/10.1016/s0190-9622\(00\)90209-3](https://doi.org/10.1016/s0190-9622(00)90209-3)
- [84] Ibbotson SH, Moseley H, Brancalion L, *et al.* Photodynamic therapy in dermatology: dundee clinical and research experience. *Photodiagnosis Photodyn Ther* 2004; 1(3): 211-223. [https://doi.org/10.1016/S1572-1000\(04\)00045-6](https://doi.org/10.1016/S1572-1000(04)00045-6)
- [85] Christie JG, Kompella UB. Ophthalmic light sensitive nanocarrier systems. *Drug Discov Today*. 2008; 13(3-4): 124-134. <https://doi.org/10.1016/j.drudis.2007.12.005>

- [86] Uchoa AF, Oliveira CS, Baptista MS. Relationship between structure and photoactivity of porphyrins derived from protoporphyrin IX. *J Porphyrins Phthalocyanines* 2010; 14: 832-845. <https://doi.org/10.1142/S108842461000263X>
- [87] Weber G, Charitat T, Baptista MS, *et al.* Lipid oxidation induces structural changes in biomimetic membranes. *Soft Matter* 2014; 10(24): 4241-4247. <https://doi.org/10.1039/c3sm52740a>
- [88] Avci P, Gupta A, Sadasivam M, *et al.* Low-level laser (light) therapy (LLLT) in skin: stimulating, healing, restoring. *Semin Cutan Med Surg* 2013; 32(1): 41-52. <https://pubmed.ncbi.nlm.nih.gov/24049929/>
- [89] Zhang P, Wu MX. A clinical review of phototherapy for psoriasis. *Lasers Med Sci* 2018; 33(1): 173-180. <https://doi.org/10.1007/s10103-017-2360-1>
- [90] Kwon HH, Lee JB, Yoon JY, *et al.* The clinical and histological effect of home-use, combination blue-red LED phototherapy for mild-to-moderate acne vulgaris in Korean patients: a double-blind, randomized controlled trial. *Br J Dermatol* 2013; 168: 1088-1094. <https://doi.org/10.1111/bjd.12186>
- [91] Niu T, Tian Y, Ren Q, Wei L, Li X, Cai Q. Red light interferes in UVA-induced photoaging of human skin fibroblast cells. *Photochem Photobiol* 2014; 90(6): 1349-1358. <https://doi.org/10.1111/php.12316>.
- [92] Wu L, Luderer M, Yang X, *et al.* Surface passivation of carbon nanoparticles with branched macromolecules influences near infrared bioimaging. *Theranostics* 2013; 3: 677-686. <https://doi.org/10.7150/thno.6535>
- [93] Kang Y-F, Li Y-H, Fang Y-W, Xu Y, Wei X-M, Yin X-B. Carbon quantum dots for zebrafish fluorescence imaging. *Sci Rep* 2015; 5: 11835. <https://doi.org/10.1038/srep11835>
- [94] Aguilar Cosme JR, Bryant HE, Claeysens F. Carbon dot-protoporphyrin IX conjugates for improved drug delivery and bioimaging. *PLoS One* 2019; 14(7): e0220210. <https://doi.org/10.1371/journal.pone.0220210>
- [95] Fischer MJ. Amine coupling through EDC/NHS: a practical approach. *Methods Mol Biol* 2010; 627: 55-73. https://doi.org/10.1007/978-1-60761-670-2_3
- [96] Zhai X, Zhang P, Liu C, *et al.* Highly luminescent carbon nanodots by microwave-assisted pyrolysis. *Chem Commun* 2012; 48: 7955-7957. <https://doi.org/10.1039/c2cc33869f>
- [97] Tsay JM, Trzoss M, Shi L, *et al.* Singlet oxygen production by Peptide-coated quantum dot-photosensitizer conjugates. *J Am Chem Soc* 2007; 129: 6865-6871. <https://doi.org/10.1021/ja070713i>
- [98] Fowley C, Nomikou N, McHale AP, McCaughan B, Callan JF. Extending the tissue penetration capability of conventional photosensitisers: a carbon quantum dot-protoporphyrin IX conjugate for use in two-photon excited photodynamic therapy. *Chem Commun (Camb)* 2013; 49: 8934-8936. <https://doi.org/10.1039/c3cc45181j>
- [99] Beack S, Kong WH, Jung HS, *et al.* Photodynamic therapy of melanoma skin cancer using carbon dot-Chlorin e6-Hyaluronate conjugate. *Acta Biomater* 2015; 26: 295-305. <https://doi.org/10.1016/j.actbio.2015.08.027>
- [100] Sun Y-P, Wang P, Lu Z, *et al.* Host-guest carbon dots for enhanced optical properties and beyond. *Sci Rep* 2015; 5: 12354 <https://doi.org/10.1038/srep12354>

- [101] Li Y, Zheng X, Zhang X, *et al.* Porphyrin-based carbon dots for photodynamic therapy of hepatoma. *Adv Healthc Mater* 2017; 6: 1600924 <https://doi.org/10.1002/adhm.201600924>
- [102] Taylor EL, Brown SB. The advantages of aminolevulinic acid photodynamic therapy in dermatology. *J Dermatolog Treat* 2002; 13(Suppl 1): S3-11. <https://doi.org/10.1080/095466302317414645>.
- [103] Smijs TG, Pavel S. The susceptibility of dermatophytes to photodynamic treatment with special focus on *Trichophyton rubrum*. *Photochem Photobiol* 2011; 87(1): 2-13. <https://doi.org/10.1111/j.1751-1097.2010.00848.x>
- [104] Calzavara-Pinton PG, Venturini M, Capezzer R, Sala R, Zane C. Photodynamic therapy of interdigital mycoses of the feet with topical application of 5-aminolevulinic acid. *Photodermatol Photoimmunol Photomed* 2004; 20(3): 144-147. <https://doi.org/10.1111/j.1600-0781.2004.00095.x>
- [105] Kamp H, Tietz HJ, Lutz M, *et al.* Antifungal effect of 5-aminolevulinic acid PDT in *Trichophyton rubrum*. *Mycoses*. 2005; 48(2): 101-107. <https://doi.org/10.1111/j.1439-0507.2004.01070.x>
- [106] Piraccini BM, Rech G, Tosti A. Photodynamic therapy of onychomycosis caused by *Trichophyton rubrum*. *J Am Acad Dermatol* 2008; 59(5 Suppl): S75-76. <https://doi.org/10.1016/j.jaad.2008.06.015>
- [107] Sotiriou E, Panagiotidou D, Ioannides D. 5-Aminolevulinic acid photodynamic therapy treatment for tinea cruris caused by *Trichophyton rubrum*: report of 10 cases. *J Eur Acad Dermatol Venereol* 2009; 23(3): 341-342. <https://doi.org/10.1111/j.1468-3083.2008.02880.x>
- [108] Sotiriou E, Koussidou T, Patsatsi A, Apalla Z, Ioannides D. 5-Aminolevulinic acid-photodynamic treatment for dermatophytic tinea pedis of interdigital type: a small clinical study. *J Eur Acad Dermatol Venereol* 2009; 23(2): 203-204. <https://doi.org/10.1111/j.1468-3083.2008.02783.x>
- [109] Prasad S, Coias J, Chen HW, Jacobe H. Utilizing UVA-1 phototherapy. *Dermatol Clin* 2020; 38(1): 79-90. <https://doi.org/10.1016/j.det.2019.08.011>
- [110] Rathod DG, Muneer H, Masood S. Phototherapy. In: *StatPearls*. Treasure Island (FL): StatPearls Publishing; 2021. Available from: <https://pubmed.ncbi.nlm.nih.gov/33085287/>
- [111] Ming L, Cheng K, Chen Y, Yang R, Chen D. Enhancement of tumor lethality of ROS in photodynamic therapy. *Cancer Med* 2021; 10(1): 257-268. <https://doi.org/10.1002/cam4.3592>
- [112] Garg AD, Krysko DV, Vandenabeele P, Agostinis P. DAMPs and PDT-mediated photo-oxidative stress: exploring the unknown. *Photochem Photobiol Sci* 2011; 10(5): 670-680. <https://doi.org/10.1039/c0pp00294a>
- [113] Rodríguez ME, Cogno IS, Milla Sanabria LS, Morán YS, Rivarola VA. Heat shock proteins in the context of photodynamic therapy: autophagy, apoptosis and immunogenic cell death. *Photochem Photobiol Sci* 2016; 15(9): 1090-1102. <https://doi.org/10.1039/c6pp00097e>
- [114] Nath S, Obaid G, Hasan T. The course of immune stimulation by photodynamic therapy: bridging fundamentals of photochemically induced immunogenic cell death to the enrichment of T-cell repertoire. *Photochem Photobiol* 2019; 95(6): 1288-1305. <https://doi.org/10.1111/php.13173>